

OB PROTEIN COMPOSITIONS AND METHOD

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Field of the Invention

5 The present invention relates to OB protein compositions and methods for preparation and use thereof.

Background

10 Although the molecular basis for obesity is largely unknown, the identification of the "OB gene" and protein encoded by ("OB protein") has shed some light on mechanisms the body uses to regulate body fat deposition. Zhang et al., Nature 372: 425-432 (1994);
15 see also, the Correction at Nature 374: 479 (1995). The OB protein has been demonstrated to be active in vivo in both ob/ob mutant mice (mice obese due to a defect in the production of the OB gene product) as well as in normal, wild type mice. The biological
20 activity manifests itself in, among other things, weight loss. To date, however, optimum conditions for obtaining the rapid weight loss in normal animals has not been ascertained. In fact, some studies have shown that, when administered by injection, rather large
25 dosages (10 mg of recombinant murine protein/kg body weight/day) are necessary for normal mice to lose 2.6% of their body weight (at the end of a 32 day period). While presently uncertain, one explanation for the necessity of such large dosages is that the optimum
30 weight loss effects are seen predominantly when the protein is in constant circulation, a condition that may not be efficiently achieved by injecting the protein.

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Summary of the Invention

The present invention stems from the observation that, as compared to administering OB protein by injection, administering OB protein by continuous pump infusion results in equivalent (or better) weight loss, in a shorter time, and with substantially lower dosages. The working example below demonstrates that a dose of 0.5 mg protein/kg body weight/day, administered via implantable osmotic pump, results in a weight loss of over 4% (as compared to baseline weight). This is in substantial contrast to other studies where similar, or less weight loss (at a comparable time point) was observed with intraperitoneal injection at the relatively high dosage of 10 mg of protein/kg body weight/day.

Thus, one aspect of the present invention is a method of treating excess weight by administering OB protein in a form for constant supply, at a dosage of less than or equal to about 1 mg protein/kg body weight/day. The dosage of less than or equal to about 1 mg protein/kg/day refers to dosages sufficient to result in observable weight loss. This is apparent from the present studies where a dosage of 0.5 mg/kg/day was sufficient to result in observable weight loss when continuously administered. In studies where injection had been the mode of administration, far higher dosages were required for weight loss. At injection dosages of 0.1 and 1 mg/kg/day, substantially no weight loss was observed in wild type (normal) mice. For example, in one study, at a comparable time point (6th day), there was a .2% loss at the 1 mg/kg dose (data not shown). Minimal weight loss was observed at the relatively high 10 mg/kg/day dose. (1.9% weight loss at day 6, data not shown). Thus, the present invention provides for dosages of 1 mg/kg/day or less

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when administered so that the supply of protein is continuous.

Connected with the present studies are the compositions and methods used for production of recombinant murine and human OB protein. The first example below discloses the preparation of recombinant murine protein, and the second example below discloses the preparation of recombinant human protein.

Additional aspects of the present invention, therefore, include the below compositions and methods for preparing recombinant murine methionyl OB protein and recombinant human methionyl OB protein, including DNA sequences, vectors, host cells, methods of fermentation, and methods of purification.

Detailed Description

The present invention stems from the observation that continuous administration of OB protein results in the need for much lower dosages for weight loss than those dosages required by acute daily injection. As set forth above, a dosage of 1 mg protein/kg body weight/day or less, continuously administered, resulted in rapid weight loss. When the underivatized protein was administered by acute injection at the 1 mg/kg/day dose, almost no weight loss in wild type (normal) mice.

The OB protein may be selected from the recombinant murine and human methionyl proteins set forth below (SEQ. ID Nos. 2 and 4) or those lacking a glutaminy residue at position 28. (See Zhang et al, Nature, supra, at page 428.) The recombinant human OB gene product is, as a mature protein, 146 amino acids; some of the DNAs obtained were observed to encode a protein lacking a glutamine residue at position 28. Zhang et al., Nature 372 at 428. The murine protein is substantially homologous to the human protein,

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particularly as a mature protein, and, further,
particularly at the N-terminus. One may prepare an
analog of the recombinant human protein by altering
(such as substituting amino acid residues), in the
5 recombinant human sequence, the amino acids which
diverge from the murine sequence. Because the
recombinant human protein has biological activity in
mice, such analog would likely be active. Proteins
lacking an N-terminal methionyl residue, such as those
10 produced by eukaryotic expression, are also available
for use.

In addition, although the present working
example involved continuous administration via
implantable pump, it is contemplated that other modes
15 of continuous administration may be practiced. For
example, chemical derivatization may result in
sustained release forms of the protein which have the
effect of continuous presence in the blood stream, in
predictable amounts. Thus, one may derivatize the
20 above proteins to effectuate such continuous
administration. The dosage of 1 mg protein/kg body
weight/day or less herein refers to the mass of
protein, exclusive of other chemical moieties used to
derivatize the protein.

25 Generally, the present protein (herein the
term "protein" is used to include "peptide", unless
otherwise indicated) may be derivatized by the
attachment of one or more chemical moieties to the
protein moiety. The chemically modified derivatives may
30 be further formulated for intraarterial,
intraperitoneal, intramuscular subcutaneous,
intravenous, oral, nasal, pulmonary, topical or other
routes of administration. Chemical modification of
biologically active proteins has been found to provide
35 additional advantages under certain circumstances, such
as increasing the stability and circulation time of the

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therapeutic protein and decreasing immunogenicity. See
U.S. Patent No. 4,179,337, Davis et al., issued
December 18, 1979. For a review, see Abuchowski et
al., in Enzymes as Drugs. (J.S. Holcerberg and J.
5 Roberts, eds. pp. 367-383 (1981)). A review article
describing protein modification and fusion proteins is
Francis, Focus on Growth Factors 3: 4-10 (May 1992)
(published by Mediscript, Mountview Court, Friern
Barnet Lane, London N20, OLD, UK). For the present
10 continuous administration, it is preferred that the
chemical modification allow for an increase in
circulation time of the protein, so that a dosage of
about 1 mg protein (exclusive of chemical moiety)/kg
body weight of a mammal/day or less will result in
15 weight loss of a mammal. The present continuous
administration will provide for weight loss of
approximately 5% of body mass in a period of 7 or fewer
days.

The chemical moieties suitable for
20 derivatization may be selected from among water soluble
polymers. The polymer selected should be water soluble
so that the protein to which it is attached does not
precipitate in an aqueous environment, such as a
physiological environment. Preferably, for therapeutic
25 use of the end-product preparation, the polymer will be
pharmaceutically acceptable. One skilled in the art
will be able to select the desired polymer based on
such considerations as whether the polymer/protein
conjugate will be used therapeutically, and if so, the
30 desired dosage, circulation time, resistance to
proteolysis, and other considerations. For the present
proteins and peptides, the effectiveness of the
derivatization may be ascertained by administering the
derivative, in the desired form (i.e., by osmotic pump,
35 or, more preferably, by injection or infusion, or,

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The polymer may be of any molecular weight, and may be branched or unbranched. For polyethylene glycol, the preferred molecular weight is between about 20 2kDa and about 100kDa (the term "about" indicating that in preparations of polyethylene glycol, some molecules will weigh more, some less, than the stated molecular weight) for ease in handling and manufacturing. Other sizes may be used, depending on the desired therapeutic 25 profile (e.g., the duration of sustained release desired, the effects, if any on biological activity, the ease in handling, the degree or lack of antigenicity and other known effects of the polyethylene glycol to a therapeutic protein or 30 analog).

The number of polymer molecules so attached may vary, and one skilled in the art will be able to ascertain the effect on function. One may mono-derivatize, or may provide for a di-, tri-, tetra- or some combination of derivatization, with the same or different chemical moieties (e.g., polymers, such as

different weights of polyethylene glycols). The proportion of polymer molecules to protein (or peptide) molecules will vary, as will their concentrations in the reaction mixture. In general, the optimum ratio (in terms of efficiency of reaction in that there is no excess unreacted protein or polymer) will be determined by factors such as the desired degree of derivatization (e.g., mono, di-, tri-, etc.), the molecular weight of the polymer selected, whether the polymer is branched or unbranched, and the reaction conditions.

The polyethylene glycol molecules (or other chemical moieties) should be attached to the protein with consideration of effects on functional or antigenic domains of the protein. There are a number of attachment methods available to those skilled in the art. E.g., EP 0 401 384 herein incorporated by reference (coupling PEG to G-CSF), see also Malik et al., Exp. Hematol. 20: 1028-1035 (1992) (reporting pegylation of GM-CSF using tresyl chloride). For example, polyethylene glycol may be covalently bound through amino acid residues via a reactive group, such as, a free amino or carboxyl group. Reactive groups are those to which an activated polyethylene glycol molecule may be bound. The amino acid residues having a free amino group may include lysine residues and the N-terminal amino acid residue. Those having a free carboxyl group may include aspartic acid residues, glutamic acid residues, and the C-terminal amino acid residue. Sulfhydryl groups may also be used as a reactive group for attaching the polyethylene glycol molecule(s). Preferred for therapeutic purposes is attachment at an amino group, such as attachment at the N-terminus or lysine group. Attachment at residues important for receptor binding should be avoided if receptor binding is desired.

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One may specifically desire N-terminally chemically modified protein. Using polyethylene glycol as an illustration of the present compositions, one may select from a variety of polyethylene glycol molecules (by molecular weight, branching, etc.), the proportion of polyethylene glycol molecules to protein (or peptide) molecules in the reaction mix, the type of pegylation reaction to be performed, and the method of obtaining the selected N-terminally pegylated protein.

The method of obtaining the N-terminally pegylated preparation (i.e., separating this moiety from other monopegylated moieties if necessary) may be by purification of the N-terminally pegylated material from a population of pegylated protein molecules.

Selective N-terminal chemical modification may be accomplished by reductive alkylation which exploits differential reactivity of different types of primary amino groups (lysine versus the N-terminal) available for derivatization in a particular protein. Under the appropriate reaction conditions, substantially selective derivatization of the protein at the N-terminus with a carbonyl group containing polymer is achieved. For example, one may selectively N-terminally pegylate the protein by performing the reaction at a pH which allows one to take advantage of the pK_a differences between the ϵ -amino group of the lysine residues and that of the α -amino group of the N-terminal residue of the protein. By such selective derivatization, attachment of a water soluble polymer to a protein is controlled: the conjugation with the polymer takes place predominantly at the N-terminus of the protein and no significant modification of other reactive groups, such as the lysine side chain amino groups, occurs. Using reductive alkylation, the water soluble polymer may be of the type described above, and should have a single reactive aldehyde for coupling to

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the protein. Polyethylene glycol propionaldehyde, containing a single reactive aldehyde, may be used.

In yet another aspect of the present invention, provided are methods of using pharmaceutical compositions of the proteins and derivatives. Such pharmaceutical compositions may be for administration for injection, or for oral, pulmonary, nasal or other forms of administration which allow for the desired circulating dose of about 1 mg protein/kg body weight/day or less. In general, comprehended by the invention are pharmaceutical compositions comprising effective amounts of protein or derivative products of the invention together with pharmaceutically acceptable diluents, preservatives, solubilizers, emulsifiers, adjuvants and/or carriers. Such compositions include diluents of various buffer content (e.g., Tris-HCl, acetate, phosphate), pH and ionic strength; additives such as detergents and solubilizing agents (e.g., Tween 80, Polysorbate 80), anti-oxidants (e.g., ascorbic acid, sodium metabisulfite), preservatives (e.g., Thimersol, benzyl alcohol) and bulking substances (e.g., lactose, mannitol); incorporation of the material into particulate preparations of polymeric compounds such as polylactic acid, polyglycolic acid, etc. or into liposomes. Hylauronic acid may also be used, and this may have the effect of promoting sustained duration in the circulation. Such compositions may influence the physical state, stability, rate of in vivo release, and rate of in vivo clearance of the present proteins and derivatives. See, e.g., Remington's Pharmaceutical Sciences, 18th Ed. (1990, Mack Publishing Co., Easton, PA 18042) pages 1435-1712 which are herein incorporated by reference. The compositions may be prepared in liquid form, or may be in dried powder, such as lyophilized form. The effective amounts are those herein described.

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The OB proteins and derivatives described are useful for modulation of the rate or quantity of fat cell deposition in a mammal. This is thought to be accomplished, in part, by a reduction in appetite, i.e., a reduction in food intake. Thus, one observable result is weight loss, or, put another way, a method of treating excess weight (via weight loss). Thus, the present compositions are useful for the manufacture of a medicament for treating excess weight in a mammal.

Another aspect is a method for reducing appetite. Either of these aspects, modulation of fat deposition or modulation of appetite, are particularly important treatments for humans (or other mammals) who desire to lose weight.

One skilled in the art will be able to ascertain other effective dosages by administration and observing weight loss. Here, the dosage of 1 mg protein/kg body weight/day or less was seen to be particularly effective, when administered on a continuous basis. More particularly, the dosage of 0.5 mg/kg body weight/day was seen to be particularly effective on normal mice. Excess weight refers to body mass for which removal is desired. It is contemplated that the present compositions and methods will be used to treat cases where removal of such excess weight (as a result of the present invention) will benefit other health concerns, such as diabetes, high blood pressure or cardiac problems, high cholesterol levels, low locomotion levels and other manifestations of excess weight. As such, the present compositions and methods may be used in conjunction with other medicaments, such as those useful for the treatment of diabetes (e.g., insulin, and possibly amylin), cholesterol and blood pressure lowering medicaments, and locomotion increasing medicaments (e.g., amphetamines). Such

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administration may be simultaneous or may be in
serriatim.

In addition, the present compositions and
methods may be used in conjunction with surgical
5 procedures, such as cosmetic surgeries designed to
alter the overall appearance of a body (e.g.,
liposuction or laser surgeries designed to reduce body
mass). The health benefits of cardiac surgeries may be
increased with concomitant use of the present
10 compositions and methods.

Therefore, the present invention encompasses
a method of treating excess weight in a mammal by
continuous administration of 1 mg protein/kg body
weight/day or less of an OB protein selected from the
15 group consisting of:

(a) recombinant methionyl murine OB protein
(~~SEQ. ID. No. 2~~);

(b) recombinant methionyl human OB protein
(~~SEQ. ID. No. 1~~);

20 (c) the protein of (a) or (b) lacking the
methionyl residue at position -1;

(d) the protein of (a), (b) or (c) lacking a
glutamine at position 28; and

(e) a chemically modified derivative of (a),
25 (b), (c) or (d), wherein the chemical modification
allows for an increase in circulation time.

Preferably, the composition of subpart (e) is
a pegylated derivative, and, more preferably, an N-
terminally pegylated derivative.

30 The derivative of subpart (e) allows for
continuous administration of the protein by increasing
the circulation time of the (unmodified) protein. The
present invention also encompasses a method of treating
excess weight where the method of continuous
35 administration is by implantable pump, such as an
osmotic pump.

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In other aspects, the present invention relates to recombinant murine and recombinant human OB DNAs and proteins, such as those of SEQ. ID NOs. 1, 2, 3, ^{4, 5, and 6} and ~~4~~, below. The recombinant proteins below are
5 bacterially expressed, and contain N-terminal methionyl residues. Vectors and host cells useful for producing such proteins are also provided. The vectors include pCFM1656 containing SEQ ID No. 1 or ~~3~~ ⁴ and host cells containing such vectors.

10 Methods for preparation of the recombinant proteins are also provided, including methods for fermentation and methods for purification.

In particular, the use of sarcosine for refolding of OB protein in solution, obtained from
15 bacterial inclusion bodies, provided for extremely efficient refolding. When proteins are expressed in bacteria, they may not be in the proper three-dimensional configuration, or, as referred to herein, properly refolded. The three dimensional configuration
20 may be critical for biological activity, and storage stability. Although Sarkosyl has been used in processes for purification of another protein (G-CSF, e.g., WO 89/10932), surprisingly, the use of sarcosine for the OB protein has resulted in a refolding
25 efficiency of over 95%. Contemplated herein is the use of N-lauroylsarcosine in a range of 0.5% - 2.0 % weight per volume of OB protein in solution (obtained from inclusion bodies). With the use of 1% sodium sarcosine, the refolding efficiency, as determined by SDS PAGE and
30 reverse phase HPLC, was 95% or greater. While one skilled in the art will recognize that other compositions may be used for refolding, the use of N-lauroyl sarcosine, as illustrated in the examples below, is particularly advantageous for providing
35 extremely efficient refolding. The removal of sarcosine was accomplished using Dowex®.

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Therefore, the present invention also includes a method of refolding partially purified OB protein in a solution obtained from inclusion bodies, said partially purified OB protein selected from the group consisting of:

(a) recombinant methionyl murine OB protein (~~SEQ. ID. No. 2~~);

(b) recombinant methionyl human OB protein (~~SEQ. ID. No. 1~~);

(c) the protein of (a) or (b) lacking the methionyl residue at position -1;

wherein said refolding is accomplished using sarcosine.

The present invention also includes methods of wherein said N-lauroyl sarcosine is used at a concentration of 0.5% - 2.0% weight per volume of solution, and, more particularly, the use of 1% N-lauroyl sarcosine. An oxidizing agent, such as copper sulfate, is also used in the refolding process.

The following examples are offered to more fully illustrate the invention, but are not to be construed as limiting the scope thereof.

EXAMPLE 1: Use of Murine OB Protein in a Continuous Pump Infusion System

This example demonstrates that continuous infusion of OB protein results in weight loss in normal mice. Normal (non-obese) mice were administered murine OB protein via osmotic pump infusion. A dosage of 0.5 mg protein/kg body weight/day resulted in a 4.62% (+/- 1.34%) loss from baseline weight by the 6th day of infusion.

MATERIALS AND METHODS

Animals: Wild type (+/+) C57B16 mice were used for this experiment. The age of the mice at the
5 initial time point was 8 weeks, and the animals were weight stabilized. 10 mice were used for each cohort (vehicle vs. protein).

Animal Handling.

10 Feeding and weight measurement. Mice were given ground rodent chow (PMI Feeds, Inc.) in powdered food
feeders (Allentown Caging and Equipment) which allowed a more accurate and sensitive measurement than use of
regular block chow. Weight was measured at the same
15 time each day (2:00 p.m.), for a period of 6 days. Body weight on the day prior to the infusion was defined as baseline weight. The mice used weighed 18-22 grams.

20 Housing. Mice were single-housed, and maintained under humane conditions.

Administration of Protein or Vehicle. Protein (as described below) or vehicle (phosphate buffered saline,
25 pH 7.4) were administered by osmotic pump infusion. Alzet osmotic minipumps (Alza, Palo Alto, CA, model no. 1007D) were surgically placed in each mice in a subcutaneous pocket in the subscapular area . The pumps were calibrated to administer 0.5 µl protein in
30 solution per hour for a dosage of 0.5 mg protein/kg body weight/day.

Controls: Control animals were those who had a Alzet osmotic minipump infusing phosphate buffered
35 saline (pH 7.4).

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Protein: Recombinant murine OB protein was used for the present experiments, generally at a concentration of about 0.9 mg/ml phosphate buffered saline, pH 7.4. The amino acid sequence (and DNA
5 sequence) used was the following:

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amino acid sequence (seq. ID. NOS. 1 and 2):

[illegible]

Herein, the first amino acid of the amino acid sequence for recombinant protein is referred to as +1, and is valine, and the amino acid at position -1 is methionine. The C-terminal amino acid is number 146

5 (cysteine).

The cloning of the murine OB DNA for expression in E. coli was done as follows. The DNA sequence was deduced from the published peptide sequence that appeared in Zhang et al., Nature 372:425-432
10 (1994). It was reverse translated using E. coli optimal codons. The terminal cloning sites were XbaI to BamHI. A ribosomal binding enhancer and a strong ribosomal binding site were included in front of the coding region. The duplex DNA sequence was synthesized using
15 standard techniques. Correct clones were confirmed by demonstrating expression of the recombinant protein and presence of the correct OB DNA sequence in the resident plasmid.

20 Expression Vector and Host Strain

The plasmid expression vector used was pCFM1656, ATCC Accession No. 69576. The above DNA was ligated into the expression vector pCFM1656 which had been linearized with XbaI and BamHI and transformed
25 into the E. coli host strain, FM5. E. coli FM5 cells were derived at Amgen Inc., Thousand Oaks, CA from E. coli K-12 strain (Bachmann, et al., Bacteriol. Rev. 40: 116-167 (1976)) and contain the integrated lambda phage repressor gene, cI₈₅₇ (Sussman et al., C.R. Acad. Sci.
30 254: 1517-1579 (1962)). Vector production, cell transformation, and colony selection were performed by standard methods. E.g., Sambrook, et al., Molecular Cloning: A Laboratory Manual, 2d Edition, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY. Host
35 cells were grown in LB media.

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Fermentation Process A three-phase fermentation protocol was used known as a fed-batch process. Media compositions are set forth below.

5 Batch: A nitrogen and phosphate source were sterilized (by raising to 122 °C for 35 minutes, 18-20 psi) in the fermentation vessel (Biolafitte, 12 liter capacity). Upon cooling, carbon, magnesium, vitamin, and trace metal sources were added aseptically. An
10 overnight culture of the above recombinant murine protein-producing bacteria (16 hours or more) of 500 mL (grown in LB broth) was added to the fermentor.

 Feed I: Upon reaching between 4.0-6.0 OD₆₀₀,
15 cultures were fed with Feed I. The glucose was fed at a limiting rate in order to control the growth rate (μ). An automated system (called the Distributive Control System) was instructed to control the growth rate to
20 0.15 generations per hour.

 Feed II: When the OD₆₀₀ had reached 30,
culture temperature was slowly increased to 42°C and the feed was changed to Feed II, below. The
fermentation was then allowed to continue for 10 hours
25 with sampling every 2 hours. After 10 hours, the contents of the fermentor was chilled to below 20°C and harvested by centrifugation.

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Media Composition:

5	Batch:	10 g/L	Yeast extract
		5.25 g/L	(NH ₄) ₂ SO ₄
		3.5 g/L	K ₂ HPO ₄
		4.0 g/L	KH ₂ PO ₄
		5.0 g/L	Glucose
		1.0 g/L	MgSO ₄ ·7H ₂ O
		2.0 mL/L	Vitamin Solution
		2.0 mL/L	Trace Metal Solution
		1.0 mL/L	P2000 Antifoam
		1.0 mL/L	P2000 Antifoam
10	Feed I:	50 g/L	Bacto-tryptone
		50 g/L	Yeast extract
		450 g/L	Glucose
		8.75 g/L	MgSO ₄ ·7H ₂ O
		10 mL/L	Vitamin Solution
15		10 mL/L	Trace Metal Solution
		10 mL/L	Trace Metal Solution
20	Feed II:	200 g/L	Bacto-tryptone
		100 g/L	Yeast extract
		110 g/L	Glucose

Vitamin Solution (Batch and Feed I):

0.5 g Biotin, 0.4 g Folic acid, and 4.2 g riboflavin, were dissolved in 450 mls H₂O and 3 mls 10 N NaOH, and brought to 500 mls in H₂O. 14 g pyridoxine-HCl and 61 g niacin were dissolved 150 ml H₂O and 50 ml 10 N NaOH, and brought to 250 ml in H₂O. 54 g pantothenic acid was dissolved in 200 ml H₂O, and brought to 250 ml. The three solutions were combined and brought to 10 liters total volume.

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Trace Metal Solution (Batch and Feed I):
Ferric Chloride ($\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$): 27 g/L
Zinc Chloride ($\text{ZnCl}_2 \cdot 4\text{H}_2\text{O}$): 2 g/L
Cobalt Chloride ($\text{CoCl}_2 \cdot 6\text{H}_2\text{O}$): 2 g/L
5 Sodium Molybdate ($\text{NaMoO}_4 \cdot 2\text{H}_2\text{O}$): 2 g/L
Calcium Chloride ($\text{CaCl}_2 \cdot 2\text{H}_2\text{O}$): 1 g/L
Cupric Sulfate ($\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$): 1.9 g/L
Boric Acid (H_3BO_3): 0.5 g/L
Manganese Chloride ($\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$): 1.6 g/L
10 Sodium Citrate dihydrate: 73.5 g/L

Purification Process for Murine OB Protein

Purification was accomplished by the following steps (unless otherwise noted, the following
15 steps were performed at 4°C):

1. Cell paste. E. coli cell paste was suspended in 5 times volume of 7 mM of EDTA, pH 7.0. The cells in the EDTA were further broken by two passes through a
20 microfluidizer. The broken cells were centrifuged at 4.2 K rpm for 1 hour in a Beckman J6-B centrifuge with a JS-4.2 rotor.

2. Inclusion body wash #1. The supernatant from above was removed, and the pellet was resuspended with
25 5 times volume of 7 mM EDTA, pH 7.0, and homogenized. This mixture was centrifuged as in step 1.

3. Inclusion body wash #2. The supernatant from above was removed, and the pellet was resuspended in ten times volume of 20 mM tris, pH 8.5, 10 mM DTT, and
30 1% deoxycholate, and homogenized. This mixture was centrifuged as in step 1.

4. Inclusion body wash #3. The supernatant from above was removed and the pellet was resuspended in ten times volume of distilled water, and homogenized. This
35 mixture was centrifuged as in step 1.

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5. Refolding. The pellet was refolded with 15 volumes of 10 mM HEPES, pH 8.5, 1% sodium sarcosine (N-lauroyl sarcosine), at room temperature. After 60 minutes, the solution is made to be 60 μ M copper sulfate, and then stirred overnight.

6. Removal of sarcosine. The refolding mixture was diluted with 5 volumes of 10 mM tris buffer, pH 7.5, and centrifuged as in step 1. The supernatant was collected, and mixed with agitation for one hour with Dowex® 1-X4 resin (Dow Chemical Co., Midland MI), 20-50 mesh, chloride form, at 0.066% total volume of diluted refolding mix. See WO 89/10932 at page 26 for more information on Dowex®. This mixture was poured into a column and the eluant was collected. Removal of sarcosine was ascertained by reverse phase HPLC.

7. Acid precipitation. The eluant from the previous step was collected, and pH adjusted to pH 5.5, and incubated for 30 minutes at room temperature. This mixture was centrifuged as in step 1.

8. Cation exchange chromatography. The pH of the supernatant from the previous step was adjusted to pH 4.2, and loaded on CM Sepharose Fast Flow (at 7% volume). 20 column volumes of salt gradient were done at 20 mM NaOAC, pH 4.2, 0 M to 1.0 M NaCl.

9. Hydrophobic interaction chromatography. The CM Sepharose pool of peak fractions (ascertained from ultraviolet absorbance) from the above step was made to be 0.2 M ammonium sulfate. A 20 column volume reverse salt gradient was done at 5 mM NaOAC, pH 4.2, with .4 M to 0 M ammonium sulfate. This material was concentrated and diafiltered into PBS.

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Results

Presented below are the percent (%) differences from baseline weight in C57Bl6J mice (8 weeks old) :

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Table 1: Weight Loss Upon Continuous Infusion

<u>Time (days)</u>	<u>Vehicle (PBS)</u>	<u>Recombinant OB protein</u>
Days 1-2	3.24 +/- 1.13	1.68 +/- 1.4
Days 3-4	4.3 +/- .97	-2.12 +/- .79
Days 5-6	4.64 +/- .96	-4.62 +/- 1.3

10 As can be seen, at the end of a 6 day continuous infusion regime, animals receiving the OB protein lost over 4% of their body weight, as compared to baseline. This is a substantially more rapid weight loss than has been observed with intraperitoneal (i.p.)
 15 injection. Weight loss at the end of a 32-day injection period, in wild type (normal) mice, with daily i.p. injections of recombinant murine OB protein at a 10 mg/kg dose was 2.6%, and had not been more than 4% at any time during the dosing schedule (data not shown).
 20 The present data indicate that with continuous infusion, a 20-fold lower dosage (0.5 mg/kg vs. 10 mg/kg) achieves more weight loss in a shorter time period.

25 The results seen here are statistically significant, e.g., -4.62% with $p < .0001$.

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EXAMPLE 2: Dose Response Studies

An additional study demonstrated that there was a dose response to continuous administration of OB protein. In this study, non-obese, CD-1 mice, weighing 35-40 g were administered recombinant murine OB protein using methods similar to the above example. The results are set forth in Table 2, below, (with % body weight lost as compared to baseline, measured as above):

Table 2: Dose Response With Continuous Administration

Dose	Time	% Reduction in body weight
0.03 mg/kg/day	Day 2	3.5
1 mg/kg/day	Day 2	7.5
1 mg/kg/day	Day 4	14

As can be seen, increasing the dose from 0.03 mg/kg/day to 1 mg/kg/day increased the weight lost from 3.5% to 7.5%. It is also noteworthy that at day 4, the 1 mg/kg/day dosage resulted in a 14% reduction in body weight.

EXAMPLE 3: Cloning and Expression of a Recombinant Human Methionyl OB Protein

This example provides compositions and methods for preparation of a recombinant human version of the OB protein.

The human version of the OB DNA was constructed from the murine OB DNA, as in Example 1, above, by replacing the region between the MluI and BamHI sites with duplex DNA (made from synthetic oligonucleotides) in which 20 codon substitutions had been designed. The MluI site is shown

under the solid line in the sequence below. This DNA
was put into the pCFM1656 vector (ATCC Accession No.
69576), in the same fashion as the recombinant murine
protein, as described above. Herein, the first amino
5 acid of the amino acid sequence for recombinant human
protein below is referred to as +1, and is valine, and
the amino acid at position -1 is methionine. The C-
terminal amino acid is number 146 (cysteine).

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Recombinant human met OB (Double Stranded) DNA and amino acid sequence (Seq. ID. Nos. *3 and 4*)

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5      1  CATATGGTACCGATCCAGAAAGTTCAGGACGACACCAAAACCTTAATTAAAACGATCGTT 60
      1  -----+-----+-----+-----+-----+-----+-----+-----+
      1  GTATACCATGGCTAGGTCTTTCAAGTCCTGCTGTGGTTTGGGAATTAATTTTGCTAGCAA
          M V P I Q K V Q D D T K T L I K T I V -

10      61  ACGCGTATCAACGACATCAGTCACACCCAGTCGGTGAGCTCTAAACAGCGTGTTACAGGC
      61  -----+-----+-----+-----+-----+-----+-----+-----+
      61  TGCGCATAGTTGCTGTAGTCAGTGTGGGTCAGCCACTCGAGATTTGTCGCACAATGTCCG
          T R I N D I S H T Q S V S S K Q R V T G -

15      121  CTGGACTTCATCCCGGGTCTGCACCCGATCCTGACCTTGTCCAAAATGGACCAGACCCCTG
      121  -----+-----+-----+-----+-----+-----+-----+-----+
      121  GACCTGAAGTAGGGCCAGACGTGGGCTAGGACTGGAACAGGTTTACCTGGTCTGGGAC
          L D F I P G L H P I L T L S K M D Q T L -

20      181  GCTGTATACCAGCAGATCTTAACCTCCATGCCGTCCCCTAACGTTCTTCAGATCTCTAAC
      181  -----+-----+-----+-----+-----+-----+-----+-----+
      181  CGACATATGGTCGTCTAGAATTGGAGGTACGGCAGGGCATTGCAAGAAGTCTAGAGATTG
          A V Y Q Q I L T S M P S R N V L Q I S N -

30      241  GACCTCGAGAACCTTCGCGACCTGCTGCACGTGCTGGCATTCTCCAAATCCTGCCACCTG
      241  -----+-----+-----+-----+-----+-----+-----+-----+
      241  CTGGAGCTCTTGGAAGCGCTGGACGACGTGCACGACCGTAAGAGGTTTAGGACGGTGGAC
          D L E N L R D L L H V L A F S K S C H L -

35      301  CCATGGGCTTCAGGTCTTGAGACTCTGGACTCTCTGGGCGGGGTCCTGGAAGCATCCGGT
      301  -----+-----+-----+-----+-----+-----+-----+-----+
      301  GGTACCCGAAGTCCAGAACTCTGAGACCTGAGAGACCCGCCCCAGGACCTTCGTAGGCCA
          P W A S G L E T L D S L G G V L E A S G -

40      361  TACAGCACCGAAGTTGTTGCTCTGTCCCGTCTGCAGGGTTCCCTTCAGGACATGCTTTGG
      361  -----+-----+-----+-----+-----+-----+-----+-----+
      361  ATGTCGTGGCTTCAACAACGAGACAGGGCAGACGTCCCAAGGGAAGTCCTGTACGAAACC
          Y S T E V V A L S R L Q G S L Q D M L W -

50      421  CAGCTGGACCTGTCTCCGGGTTGTTAATGGATCC
      421  -----+-----+-----+-----+-----+-----+-----+-----+
      421  GTCGACCTGGACAGAGGCCCAACAATTACCTAGG
          Q L D L S P G C *

55

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Fermentation: Fermentation of the above host cells to produce recombinant human OB protein was accomplished using the conditions and compositions as described above

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for recombinant murine material. The results were analyzed for yield (grams ob DNA product/liter of fermentation broth), prior to purification of the recombinant human OB material. (Minor amounts of bacterial protein were present.) Bacterial expression was also calculated.

Table 3: Analysis of Human OB Protein Expression

Timepoint	OD (@600 nm)	Yield (g/L)	Expression (mg/OD·L)
Ind. + 2 hours.	47	1.91	41
Ind. + 4 hours.	79	9.48	120
Ind. + 6 hours.	95	13.01	137
Ind. + 8 hours.	94	13.24	141
Ind. + 10 hours.	98	14.65	149

abbreviations: Ind. + __ hours means the hours after induction of protein expression, as described in Example I for the recombinant murine material using

pCFM1656

OD: optical density, as measured by spectrophotometer
milligrams per OD unit per liter

mg/OD·L: expression in terms of milligrams of protein per OD unit per liter.

g/L: grams protein/liter fermentation broth

Purification of the recombinant human OB protein: Recombinant human protein may be purified using methods similar to those used for purification of recombinant murine protein, as in Example 1, above. For preparation of recombinant human OB protein, step 8 was performed by adjusting the pH of the supernatant from step 7 to pH 5.0, and loading this onto a CM Sepharose fast flow column. The 20 column volume salt gradient was performed at 20 mM NaOAC, pH 5.5, 0M to 0.5 M NaCl. Step 9 was performed by diluting the CM Sepharose pool four fold with water, and adjusting the pH to 7.5. This mixture was made to 0.7 M ammonium sulfate. Twenty column volume reverse salt gradient was done at 5 mM NaOAC, pH 5.5, 0.2 M to 0M ammonium sulfate. Otherwise, the above steps were identical.

While the present invention has been described in terms of preferred embodiments, it is understood that variations and modifications will occur to those skilled in the art. Therefore, it is intended that the appended claims cover all such equivalent variations which come within the scope of the invention as claimed.

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